SUPEROXIDE REDUCTION AS A MECHANISM OF ASCORBATE-STIMULATED OXYGEN UPTAKE BY ISOLATED CHLOROPLASTS

J.F. Allen and D.O. Hall

University of London, King's College, Botany Department, 68 Half Moon Lane, London SE24 9JF

Received April 16,1973

SUMMARY: Addition of lmM ascorbate to isolated chloroplasts with methyl viologen (MV) as electron acceptor trebled the rate of oxygen uptake and decreased the ADP/O ratio to a third of that with no ascorbate present. These effects of ascorbate were reversed by superoxide dismutase (SOD), which in the absence of ascorbate had little effect on O_2 uptake or ADP/O ratio. A chloroplast-associated SOD activity equivalent to 500 units/mg chlorophyll was detected. The effects of ascorbate and SOD on O_2 uptake were similar in both coupled and uncoupled chloroplasts. The results are consistent with the hypothesis that ascorbate stimulates O_2 uptake by reduction of superoxide, which is formed by autoxidation of the added electron acceptor (MV), and which dismutates in the absence of ascorbate. Ascorbate does not seem to stimulate O_2 uptake by replacing water as the photosystem II donor.

Ascorbate has been reported to act as a photosystem (PS) II donor in chloroplasts with water-oxidation inactivated by heating^{1,2} or tristreatment³. Bohme and Trebst² have suggested that ascorbate-stimulated O₂ uptake by isolated chloroplasts - which are still able to perform the Hill reaction in the absence of ascorbate - may be explained by ascorbate replacing water as the PSII electron donor. They suggest that electron transport in coupled chloroplasts is accelerated by ascorbate, since electron donation by ascorbate bypasses a normally rate-limiting PSII phosphorylation site.

Epel and Neumann⁴ have, however, proposed an alternative mechanism of ascorbate-stimulated O_2 uptake in the presence of a low potential PSI acceptor. In this scheme superoxide, produced by autoxidation of the acceptor, is reduced by ascorbate to peroxide (as has also been proposed by Elstner et al⁵), rather than the superoxide dismutating to peroxide and oxygen, which occurs in the absence of ascorbate. Aerobic oxidation of the semiquinone of low potential dyes, such as methyl viologen, has been

reported to involve superoxide formation^{6,7}.

The results reported here support Epel and Neumann's scheme in three ways: 1) the observed ascorbate stimulation of O_2 uptake and depression of the ADP/O ratio require only that ascorbate trebles the net O_2 uptake per pair of electrons transferred from water, by reducing the superoxide formed to H_2O_2 rather than the superoxide dismutating to H_2O_2 and oxygen; 2) the addition of superoxide dismutase reverses the effects of ascorbate; 3) the effects of ascorbate and SOD on O_2 uptake are similar for both coupled and NH₄Cl-uncoupled chloroplasts.

Chloroplasts (unbroken; type B of Hall's classification 8) were METHODS: isolated from greenhouse spinach by a method based on that of Hall et al9, using a Polytron homogenizer, with ascorbate omitted from the grinding medium. Broken, washed chloroplasts (type C) were prepared by resuspending the chloroplasts in 50 mls of tenfold diluted resuspending medium; centrifuging for 4 minutes at 4000g and resuspending in undiluted medium. Chlorophyll estimation was performed as described by Arnon 10. 0, uptake was measured in a Rank O2 electrode. Illumination by two 300W slide projectors gave an intensity of 8.8 x 104 erg cm⁻² sec⁻¹, with Cinemoid 5A filters transmitting light of wavelengths between 540 nm and 740 nm. ADP/O ratios were calculated by the method of Hall et al 9. SOD, isolated as bovine erythrocuprein by the method of Weser et all, was kindly supplied by U. Weser and was added to the reaction vessel at 8000 units/ml. RESULTS AND DISCUSSION: Addition of ascorbate to both coupled and NH4Cluncoupled broken, washed (type C) chloroplasts gave a three-fold stimulation of O2 uptake at concentrations greater than O.5mM (Figure 1). With unbroken type B chloroplasts (which were swollen in the hypotonic reaction medium) maximal ascorbate stimulation was only 250% of the rate without ascorbate, but there was still no significant difference between ascorbate stimulation

of the coupled and uncoupled rates.

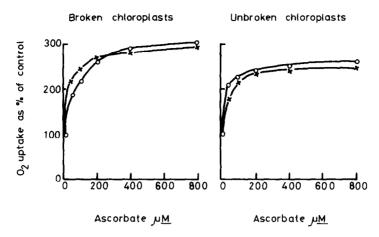


Figure 1 O₂ uptake as % of the minus-ascorbate rate versus ascorbate concentration in the reaction medium, which also contained 0.1M sorbitol, 5mM MgCl₂, 20mM NaCl, 2mM EDTA, 50mM HEPES pH 7.5, 3mM NaN₃, 50μ M MV, and chloroplasts equivalent to 100 μ g of chlorophyll, in a final volume of 2 ml. • coupled rate. × ; uncoupled rate (+5mM NH₄Cl).

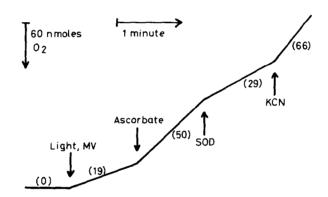
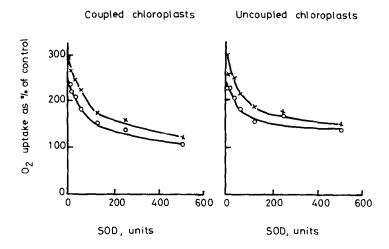


Figure 2 The effects of ascorbate (lmM), SOD (400 units), and KCN (l0mM) on O_2 uptake by broken chloroplasts in the electrode. Other conditions as in Figure 1. Bracketed figures are the rates of O_2 uptake in μ moles/mg chl/hr.

The inhibitory effect of SOD was completely reversed by 10mM KCN, which caused up to a trebling of the SOD-inhibited rate, and which caused only a 20% stimulation of the rate of O_2 uptake in the absence of SOD (T_a ble 1). The effects on chloroplast O_2 uptake of addition of ascorbate, SOD and KCN, are shown in the trace reproduced in Figure 2.

The effect of SOD on O_2 uptake stimulated by $lm\underline{M}$ ascorbate is shown for coupled and uncoupled chloroplasts in Figure 3. In both cases the



difference between unbroken (type B) and broken (type C) chloroplasts corresponds to a chloroplast-associated SOD activity of about 50 units in chloroplasts containing 100 µg of chlorophyll. From this one can calculate that there are 5-10 SOD molecules per electron transport chain.

Ascorbate and SOD had effects on the ADP/O ratios of both unbroken and broken, washed chloroplasts (Table 1), analogous to their effects on O2 uptake. Figure 4 shows the ADP/O ratio of the chloroplasts plotted against the concentration of ascorbate in the reaction medium. For broken chloroplasts, the ADP/O ratio is depressed by ascorbate to about a third of its original value, and to slightly more than a third for unbroken chloroplasts. If the exygen taken up per electron pair is trebled by the addition of ascorbate, as suggested by Figure 1, the decrease in the observed ADP/O ratio (Figure 4) does not require the assumption that the ATP/2e has decreased. Hence these results do not require that electrons from ascorbate pass through only one of the two phosphorylation sites that are now thought to be involved in non-cyclic photophosphorylation 9,12-16.

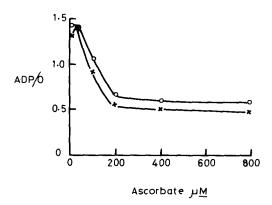


Figure 4 ADP/O versus ascorbate concentration in reaction medium. Conditions as for Figure 1, with 10mM K₂HPO₄ and 0.125 mM ADP added. o—o; unbroken (type B) chloroplasts. x—x; broken, (type C) chloroplasts.

Table 1 The effects of ascorbate (lmM), SOD (400 units), and KCN (lomM) on coupled O_2 uptake and the ADP/O ratio for broken (type C) chloroplasts. Reaction conditions as in Figure 4, i.e. all rates are with methyl viologen as electron acceptor.

Additions	O ₂ uptake µmoles/mg chl/hr	ADP/O
None	33	1.44
KCN alone	38	1.49
SOD alone	29	1.67
Ascorbate alone	86	0.63
Ascorbate + KCN	84	0.67
Ascorbate + SOD	20	1.44
Ascorbate + SOD + KCN	7 2	0.77

Photosynthetic transport of a pair of electrons from water to a low potential dye such as methyl viologen is summarized in equation 1. Aerobic autoxidation of the dye may produce superoxide ions, (equation 2), and spontaneous superoxide dismutation may then occur (equation 3). If catalase is inhibited by azide so that no breakdown of $\mathbf{H}_{\mathbf{g}}\mathbf{O}_{2}$ occurs, the net \mathbf{O}_{2} uptake per pair of electrons $(\mathbf{O}_{2}\phi/2e^{-})$ may be deduced from the summation of equations 1-3 to give equation 4, the Mehler reaction:

$$H_{2}O + 2MV_{ox} \longrightarrow \frac{1}{2}O_{2}\uparrow + 2 MV_{red} + 2H^{+} \dots 1$$

$$2MV_{red} + 2O_{2} \downarrow \longrightarrow 2 MV_{ox} + 2O_{2}^{-} \dots 2$$

$$2O_{2}^{-} + 2H^{+} \longrightarrow H_{2}O_{2} + O_{2}\uparrow \dots 3$$

$$H_{2}O + \frac{1}{2}O_{2} \downarrow \longrightarrow H_{2}O_{2} \dots 4$$

Hence $0_2 \sqrt[4]{2e^-} = \frac{1}{2}$ in this system; i.e. 0_2 uptake proceeds at the same rate as 0_2 evolution in a Hill reaction with ferricyanide.

If ascorbate replaces water as the PSII donor, producing dehydroascorbate (DHA), equation 5 replaces equation 1, and the net reaction becomes that shown in equation 6, the same as that proposed by Elstner et al⁵

In this case $0_2 \psi/2e^- = 1$. Hence the effect of ascorbate addition would be to double the rate of 0_2 uptake for any given rate of electron transport, as proposed by Bohme and Trebst².

If, however, ascorbate exerts its effect by reducing to $\rm H_2O_2$ the superoxide formed in reaction 2 (as suggested by Epel and Neumann⁴), equation 7 replaces equation 3, and the overall reaction becomes that expressed in equation 8.

Here $0_2 \oint /2e^- = 1\frac{1}{2}$; thus the ratio is trebled (from $\frac{1}{2}$ to $1\frac{1}{2}$) by the addition of ascorbate. SOD catalyzes reaction 3 and may interact with 0_2 before ascorbate can, so the observed reversal of ascorbate's effect is predicted by the scheme summarized in equation 8.

The data presented here also show the predicted trebling of the rate of O_2 uptake on addition of ascorbate (Figure 1), which occurs in both coupled and uncoupled chloroplasts. Since the trebling occurs in uncoupled chloroplasts it cannot be a result of electrons from ascorbate by-passing a ratelimiting PSII phosphorylation site². Bohme and Trebst² report a constant rate of ATP formation in chloroplasts with and without ascorbate, which in

our view may be adequately explained by an unchanged rate of electron transport through both phosphorylation sites. This conclusion is supported by the reversal by added SOD of ascorbate-depression of the ADP/O ratio (Table 1) and of ascorbate-stimulation of O2 uptake (Figure 3).

The mechanism of ascorbate-stimulated O2 uptake, proposed by Epel and Neumann4, has the added advantage that it predicts an absence of stimulation by ascorbate of NADP reduction in untreated chloroplasts 2. since it does not require the ad hoc assumption that the effectiveness of a PSII electron donor depends directly on the chemical nature of the terminal acceptor.

Thanks are due to Dr. S.G. Reeves, Dr. M.C.W. Evans and Mr. J. Lumsden for helpful discussion. JFA is in receipt of a Science Research Council Research Studentship.

REFERENCES:

- Katoh, S. and San Pietro, A., Arch. Biochem. Biophys. 122, 144 (1967). 1.
- Böhme, H. and Trebst, A., Biochim. Biophys. Acta, 180, 137 (1969). 2.
- 3.
- Yamashita, T. and Butler, W.L., Plant Physiol. 44, 435 (1969). Epel, B.L. and Neumann, J., VI. Intl. Congr. Photobiology, 4. Bochum, Germany. Abstr. No. 237 (1972).
- Elstmer, E.F., Heupel, A. and Vaklinova, S., Z. Pflanzenphysiol, 5• <u>62</u>, 184 (1970).
- 6. McCord, J.M., and Fridovitch, I., J. Biol. Chem. 245, 1374 (1970).
- Stancliffe, T.C. and Pirie, A., FEBS Letters 17, 297 (1971). 7.
- Hall, D.O., Nature New Biol. 235, 125 (1972). 8.
- Hall, D.O., Reeves, S.G. and Baltscheffsky, H., Biochem. 9. Biophys. Res. Commun. 43, 359 (1971).
- 10. Arnon, D.I., Plant Physiol. 24, 1 (1949).
- Weser, U., Bunnenberg, E., Cammack, R., Djerassi, C., Flohe, L., 11. Thomas, G., and Voelter, W., Biochim. Biophys. Acta, 243, 203 (1971)
- 12. Izawa, S. and Good, N.E., Biochim. Biophys. Acta. 162, 380 (1968).
- Horton, A.A. and Hall, D.O., Nature, 218, 386 (1968). 13.
- West, K.R. and Wiskich, J.T., Biochem. J. 109, 527 (1968). Reeves, S.G., Heathcote, P. and Hall, D.O., VI. Intl. Congr. Photobiology, 15. Bochum, Germany. Abstr. No. 284 (1972). Also: Reeves, S.G., Ph.D. Thesis, King's College London (1972).
- 16. West, K.R. and Wiskich, J.T., Biochim. Biophys. Acta, 292, 197 (1973).